

NOTES

AB-400, a New Tetraene Macrolide Isolated from *Streptomyces costae*

LIBRADA M. CAÑEDO^{*†}, LUIS COSTA, LUIS M. CRIADO^{††},
JOSÉ L. FERNÁNDEZ PUENTES[†] and MIGUEL A. MORENO

Departamento de Investigación, Antibióticos S.A.,
24080 León, Spain

KENNETH L. RINEHART

Roger Adams Laboratory, University of Illinois,
Urbana, Illinois 61801, U.S.A.

(Received for publication February 21, 2000)

In our screening program for new antifungal compounds produced by microorganisms, a new natural tetraene macrolide AB-400 has been isolated from culture broth of *Streptomyces costae*. In addition, this strain produces the known tetraene antibiotic pimaricin. Both antibiotics were found to have very similar structure and were compared by physico-chemical, spectral and microbiological methods.

The compound suppressed the growth of fungi and the activity is compared with another polyene macrolide, amphotericin B, nystatin, candicidin and pimaricin. The structure of AB-400 and its *N*-acetyl derivative (Fig. 1) were determined by comparative physico-chemical and spectral analysis with pimaricin and *N*-acetylpimaricin.

Experimental

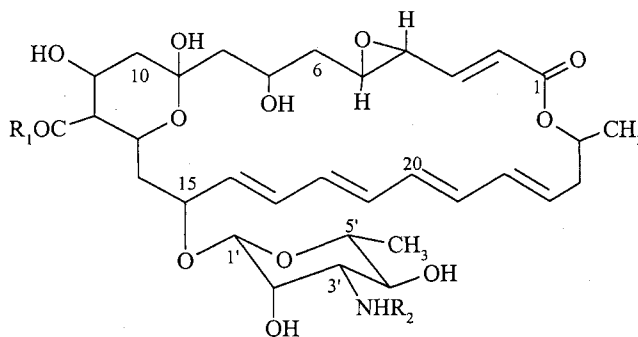
The microorganism was isolated from a soil sample collected in the surrounding of Madrid, Spain. Taxonomic studies indicated that it belonged to the genus *Streptomyces*.

A suspension of spores of the microorganism was inoculated into 1 liter Erlenmeyer flask containing 200 ml of a seed medium composed of glucose 5%, cotton seed meal 2%, corn steep liquor 1%, dextrin 1% and CaCO₃ 0.1%. The pH was adjusted to 6.2 before sterilization and

incubated at 28°C for 5 days on an orbital shaking table. The thus prepared inoculum was used to seed 8-liter jar fermenter containing 4 liters of the production medium composed of soybean meal 4%, glucose 6%, soybean oil 0.5% and CaCO₃ 1%. The pH was adjusted to 6.2 before sterilization. The fermentation was carried out at 28°C for 90 hours, after this time the amount of the antibiotic has reached its maximum and the culture broth was harvested. The production of the antibiotic during the fermentation and the following extraction and purification processes were performed by HPLC. Spherisorb ODS-2 C18 (Waters, 10 μm) column was used for HPLC analysis. The solvent system, acetonitrile-0.06 M citrate buffer (3:7, pH 4.8), was used as the mobile phase. The flow rate was 1.5 ml/minute and the monitored wavelength was 305 nm. HPLC analysis gave the following retention times (minute): 4.5 (2), 6.6 (1), 4.1 (4) and 6.1 (3).

The fermentation broth (2 liters) was adjusted to pH 8 with 5 N NaOH and it was filtrated to separate mycelium and supernatant. The micelial cake was extracted twice with 400 ml of 2-propanol, the extract was concentrated *in vacuo* to 200 ml and was diluted with 200 ml of H₂O. The

Fig. 1. The structures of AB-400 (1), pimaricin (2) and their *N*-acetyl derivatives (3) and (4).



AB-400 (1)	R ₁ = NH ₂	R ₂ = H
Pimaricin (2)	R ₁ = OH	R ₂ = H
<i>N</i> -acetyl AB-400 (3)	R ₁ = NH ₂	R ₂ = COCH ₃
<i>N</i> -acetylpimaricin (4)	R ₁ = OH	R ₂ = COCH ₃

[†] Present address: Instituto Biomar, S.A. Edificio CEI, Onzonilla, 24231 León, Spain.

^{††} Present address: Centro Nacional de Biotecnología, Universidad Autónoma de Madrid, 28049 Madrid, Spain.

Table 1. Comparison of ^{13}C NMR chemical shifts (δ , ppm) for **3** and **4** (125 MHz, pyridine- d_5).

Carbon No.	3	4
1	165.2	165.2
2	125.3	125.2
3	145.2	145.1
4	54.6	54.5
5	59.2	59.1
6	41.9	41.8
7	67.6	67.6
8	47.8	47.7
9	98.3	98.4
10	39.7	39.6
11	66.6	66.8
12	59.5	59.4
13	66.3	66.5
14	37.9	38.3
15	75.4	75.5
16	134.2	134.0
17	129.1	129.3
18	131.8	132.0
19	132.1	132.2
20	132.6	132.5
21	136.4	136.3
22	137.6	137.8
23	128.8	128.8
24	45.9	45.7
25	70.1	70.0
26	20.4	20.3
27	176.7	176.8
1'	98.0	98.1
2'	71.0	71.0
3'	56.4	56.4
4'	72.6	72.7
5'	74.8	74.6
6'	18.9	18.5
3'-NHCOCH ₃		
CH ₃ CO	171.4	171.2
CH ₃ CO	23.2	23.1

Assignments to the numbering system shown in Fig. 1.

resultant aqueous alcohol extract was adjusted to pH 10 with NaOH and it was stood one overnight at -5°C . The precipitate was filtered and washed with water to yield 1.2 g of AB-400 crude powder. Pimaricin was concentrated in the supernatant of that precipitation. The crude of AB-400 was chromatographed on a silicagel column packed with CHCl_3 . The column after washing with CHCl_3 -MeOH 1:1 was eluted with MeOH. The eluate was monitored by HPLC and the fractions containing AB-400 as a peak by HPLC were collected and concentrated to dryness in the dark to yield 125 mg of fine yellow powder. To enhance the

solubility of the antibiotic, AB-400 was derivated like *N*-acetyl derivative and purified by FLC on silica gel using CHCl_3 -MeOH 75:25 as eluting solvent.

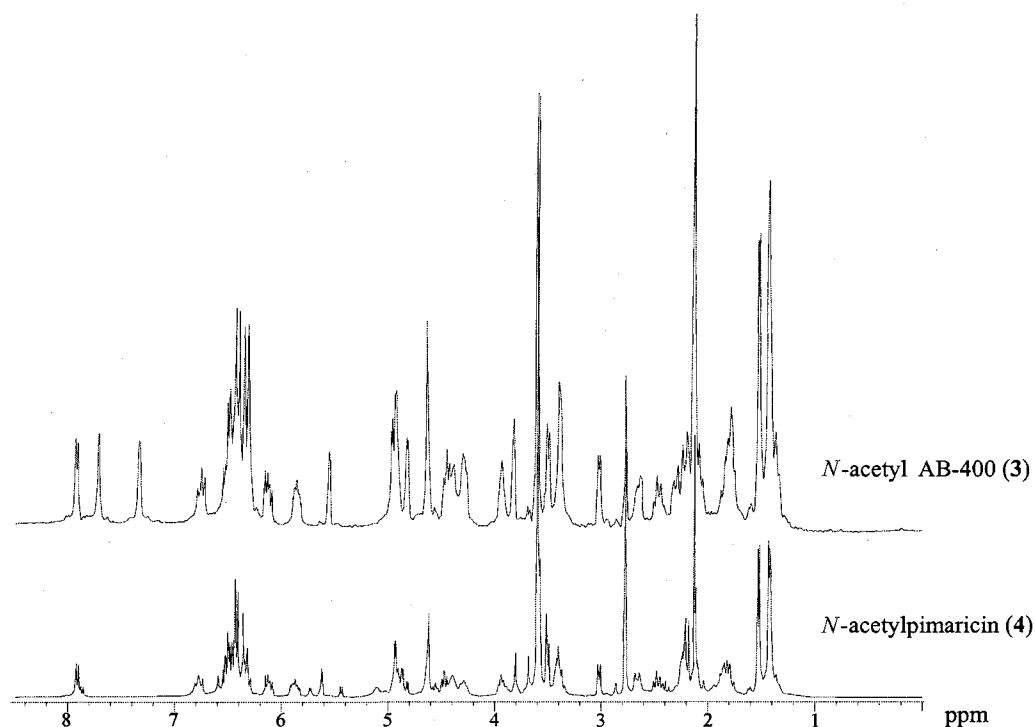
Results and Discussion

AB-400 and pimaricin were transformed into their *N*-acetyl derivatives (**3** and **4**, Fig. 1) by the reported method¹⁾, which facilitated the purification process. The used pimaricin was a commercially available authentic sample from Sigma Chem. Com. Pimaricin (**2**) and its *N*-acetyl derivative (**4**) were identified by comparison of their NMR and Mass spectra with data given as reference²⁻⁵⁾. AB-400 (**1**) and *N*-acetylAB-400 (**3**), exhibited the physico-chemical properties similar to pimaricin (**2**) and *N*-acetylpimaricin (**4**), these components are distinguished from each other by their HPLC mobilities. The UV spectrum of **1** showed characteristic polyene-type absorption very similar to that of **2** with maxima at 290, 304, and 318 nm corresponding to an all-*trans*-tetraene moiety⁶⁾. The IR spectrum of **1**, ν_{max} (KBr) cm^{-1} : 3448, 1068 indicated the presence of hydroxyl group, a lactone unit at 1721, double bonds at 1630, there is no band at 1579 of an ionized carboxyl unit and it is shown a new band at 1655 of an amide group. The molecular formula of **1** $\text{C}_{33}\text{H}_{48}\text{N}_2\text{O}_{12}$ was determined by HRFAB-MS experiment, which gave a $(\text{M}+\text{H})^+$ ion at m/z 665.3280 (calcd. m/z 665.3286 for $\text{C}_{33}\text{H}_{49}\text{N}_2\text{O}_{12}$).

The molecular formula of **3** was determined to be $\text{C}_{35}\text{H}_{50}\text{N}_2\text{O}_{13}$ from the FAB-MS peaks at m/z 706.9 $(\text{M}+\text{H})^+$ and m/z 705.3 $(\text{M}-\text{H})^-$ and m/z 687.5 $(\text{M}-\text{H}-\text{H}_2\text{O})^-$ and from HRFAB-MS [found m/z 705.3225 $(\text{M}-\text{H})^-$, calcd. m/z 705.3235 for $\text{C}_{35}\text{H}_{49}\text{N}_2\text{O}_{13}$], which was supported by the number of carbons in the ^{13}C NMR spectrum (Table 1). The molecular formula of **4** was determined to be $\text{C}_{35}\text{H}_{49}\text{NO}_{14}$ from the FAB-MS peaks at m/z 707.9 $(\text{M}+\text{H})^+$ and m/z 706.2 $(\text{M}-\text{H})^-$ and m/z 688.4 $(\text{M}-\text{H}-\text{H}_2\text{O})^-$ and from HRFAB-MS [found m/z 706.3094 $(\text{M}-\text{H})^-$, calcd. m/z 706.3075 for $\text{C}_{35}\text{H}_{48}\text{NO}_{14}$]. The molecular weight of both compounds differs in one mass unit. The molecular formula of **3** shows one nitrogen and one hydrogen plus than **4** and one oxygen less than **4**.

The ^{13}C NMR spectra of **3** and **4** recorded in pyridine- d_5 solution are virtually superimposable (Table 1). HMQC experiments allowed the specific ^{13}C assignments and multiplicities were determined by DEPT experiments. Assignments in ^{13}C NMR chemical shifts of **3** and **4** are compared in Table 1.

The ^1H NMR spectra of **3** and **4** were almost identical when the spectra were recorded in pyridine- d_5 . However,

Fig. 2. ^1H NMR spectra of **3** and **4** in $\text{DMSO-}d_6$ (500 MHz).

when the spectra were recorded in $\text{DMSO-}d_6$, a striking difference can be noticed in the behavior at 7~8 ppm (Fig. 2). The ^1H NMR of **3** shows signals of amide group, two broad singlets at 7.28 and 7.68 ppm which disappear by adding NaOD. In the ^1H NMR spectrum of **4** no signals of the amide protons could be detected (Fig. 2). The proton correlations from $^1\text{H-}^1\text{H}$ COSY support the structure of **3** and were very similar to those observed on the $^1\text{H-}^1\text{H}$ COSY of **4**, the only difference was that the COSY of **3** showed that the protons at 7.28 and 7.68 ppm were coupled. The structure assignment of **3** is consistent with a compound with amide group in place of the corresponding carboxylic group in **4** as shown in Fig. 1. This assignment was confirmed by methylation experiments of **3** and **4**, the methyl esters were prepared by the reported method⁷⁾. The compound **4** was treated with CH_2N_2 in THF and the methyl ester was obtained which gave a clear molecular ion $(\text{M-H})^-$ at m/z 720.3. In the case of **3**, the methyl ester was not yielded in the same conditions. For all of these findings, the total planar structure of **1** was established as shown in Fig. 1.

Biological Activity

In vitro antifungal activity of AB-400 and *N*-acetyl AB-400 was tested against *Candida albicans* and *Candida tropicalis*, in comparison with other polyene antibiotics such as nystatin, amphotericin B, candicidin and pimaricin. The results obtained with the antibiotics in the serial dilution test⁸⁾ are compared in Table 2. AB-400 was active against the organisms tested with potency comparable to those of other tetraene antibiotics such as nystatin and pimaricin. Acylation of AB-400 drastically reduces the activity.

Acknowledgments

The authors are grateful to Mr. FURONG SUN, University of Illinois, Urbana, for mass spectroscopy. We wish to thank Drs. A. SAN FELICIANO and M. MEDARDE, School of Pharmaceutical Science, Salamanca University, for measurement of NMR spectra.

Table 2. Antifungal activity of AB-400, *N*-acetyl AB-400 and other polyene macrolides (MIC, μM).

STRAIN	AB-400	<i>N</i> -acetyl AB-400	AMPH-B	CAND	NYS	PIM
<i>Candida albicans</i>	18.0	>75	0.86	0.21	3.3	18.0
<i>Candida tropicalis</i>	4.6	>75	1.7	0.43	3.3	9.3

AMPH-B: amphotericin B, CAND: candicidin, PIM: pimaricin, NYS: nystatin

References

- 1) MECHLINSKI, W. & C. P. SCHAFFNER: Polyene macrolide derivatives. I. *N*-Acylation and esterification reactions with amphotericin B. *J. Antibiotics* 25: 256~258, 1972
- 2) PANDEY, R. C. & K. L. RINEHART: Carbon-13 nuclear magnetic resonance evidence for cyclic hemiketals in the polyene antibiotics amphotericin B, nystatin A₁, tetrin A, tetrin B, lucensomycin and pimaricin. *J. Antibiotics* 29: 1035~1042, 1976
- 3) CEDER, O.; B. HANSSON & U. RAPP: Pimaricin-VIII. Structural and configurational studies by electron impact and field desorption mass spectrometry, ¹³C (25.2 MHz) and ¹H (279 MHz)-NMR spectroscopy. *Tetrahedron* 33: 2703~2714, 1977
- 4) RADICS, L.; M. INCZE, K. DORNBERGER & H. THRUM: Tetramycin B, a new polyene macrolide antibiotic. The structure of tetramycins A and B as studied by high-field NMR spectroscopy. *Tetrahedron* 38: 183~189, 1982
- 5) LANCELIN, J. M. & J. M. BEAU: Stereostructure of pimaricin. *J. Am. Chem. Soc.* 112: 4060~4061, 1990
- 6) HIROTA, H.; A. ITOH, J. IDO, Y. IWAMOTO, E. GOSHIMA, T. MIKI, K. HASUDA & Y. OHASHI: YS-822A, a new polyene macrolide antibiotic. II. Planar structure of YS-822A. *J. Antibiotics* 44: 181~186, 1991
- 7) PANDEY, R. C. & K. L. RINEHART: An improved method for the preparation of methyl esters of polyene antibiotics. *J. Antibiotics* 30: 158~162, 1977
- 8) RADETSKY, M.; R. C. WHEELER, M. H. ROE & J. K. TODD: Microtiter broth dilution method for yeast susceptibility testing with validation by clinical outcome. *J. Clin. Microbiol.* 24: 600~606, 1986